



Three new species of *Aleurodiscus* s.l. (Russulales, Basidiomycota) from southern China

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Abstract

Three new species of *Aleurodiscus* s.l. with corticioid basidiomata are described and illustrated from southern China based on morphological evidence and phylogenetic analyses of ITS and nrLSU sequence data. *Aleurodiscus bambusinus* was collected from Jiangxi Province on bamboo and is distinct by having a compact texture, simple-septate generative hyphae, abundant acanthophyses, basidia with acanthophysoid appendages and smooth basidiospores. *Aleurodiscus isabellinus* was collected from Yunnan Province on both angiosperm wood and bamboo and is distinct by having soft basidiomata with yellow to yellowish-brown hymenophore, yellow acanthophyses, simple-septate generative hyphae and smooth basidiospores. *Aleurodiscus subroseus* was collected from Guangxi Autonomous Region and Guizhou Province on angiosperm wood and is distinct by having pinkish basidiomata when fresh, clamped generative hyphae, clavate acanthophyses and echinulate basidiospores. In the phylogenetic tree, *A. bambusinus* and *A. isabellinus* were nested within the *A. cerussatus* group, whilst *A. subroseus* was clustered with *A. wakefieldiae*. An identification key to 26 species of *Aleurodiscus* s.l. in China is provided.

Keywords

acanthophyses, corticioid fungi, Stereaceae, taxonomy, wood-inhabiting fungi

Introduction

Aleurodiscus s.l. is a large group of wood-inhabiting fungi with a broad morphological circumscription. It is characterised by having cupulate, effused or effused-reflexed basidiomata, a monomitic or dimitic hyphal system with simple-septate or clamped gen-

erative hyphae, smooth or ornamented, amyloid basidiospores and sterile organs such as acanthophyses, gloeocystidia and dendrohyphidia (Núñez and Ryvarden 1997). Although *Aleurodiscus* s.l. had been divided into several small genera based on different combinations of morphological characters, phylogenetic analyses did not fully support these separations (Wu et al. 2001; Dai and He 2016). Accordingly, the inter- and intrageneric phylogeny of *Aleurodiscus* s.l. in Stereaceae is still unclear and no reliable morphological characters can be used to recognise the small segregated genera. Thus, the broad sense concept of the genus has often been adopted by mycologists when describing new species (Núñez and Ryvarden 1997; Gorjón et al. 2013; Dai et al. 2017a, b).

A recent survey on *Aleurodiscus* s.l. from China (Dai and He 2016, 2017, Dai et al. 2017a, b) revealed that its species diversity is high and many species, especially those with corticioid basidiomata on both herbaceous and ligneous plants, are still undescribed. In the present study, three new species are described and illustrated from southern China, amongst which two species have abundant acanthophyses and smooth basidiospores and one species bears echinulate basidiospores. Morphological differences between new species and their relatives are discussed. Their phylogenetic positions were inferred from a combined dataset of ITS and nrLSU sequence data.

Materials and methods

Morphological studies

Voucher specimens are deposited in the herbaria of Beijing Forestry University, Beijing, China (BJFC), Centre for Forest Mycology Research, U.S. Forest Service, Madison, USA (CFMR) and Southwest Forestry University, Kunming, China (SWFC). Freehand sections were made from basidiomata and mounted in 2% (w/v) potassium hydroxide (KOH), 1% phloxine (w/v) or Melzer's reagent. Microscopic examinations were carried out with a Nikon Eclipse 80i microscope at magnifications up to $1000 \times$. Drawings were made with the aid of a drawing tube. The following abbreviations are used: L = mean spore length, W = mean spore width, Q = L/W ratio, n (a/b) = number of spores (a) measured from number of specimens (b). Colour names and codes follow Kornerup and Wanscher (1978).

DNA extraction and sequencing

A CTAB plant genome rapid extraction kit-DN14 (Aidlab Biotechnologies Co. Ltd, Beijing) was employed for DNA extraction and PCR amplification from dried specimens. The ITS and nrLSU gene regions were amplified with primer pairs ITS5/ITS4 (White et al. 1990) and LR0R/LR7 (http://www.biology.duke.edu/fungi/mycolab/primers.htm), respectively. The PCR procedures followed Dai and He (2016). DNA sequencing was performed at Beijing Genomics Institute and the sequences were deposited in GenBank.

Phylogenetic analyses

The molecular phylogeny was inferred from a combined dataset of ITS and nrLSU sequences of representative members of Stereaceae sensu Larsson (2007) (Table 1). The ingroup taxa sampling and outgroup selection followed Dai et al. (2017b). The sequences were aligned using MAFFT v.6 (Katoh and Toh 2008, http://mafft.cbrc.jp/alignment/server/). Alignments were optimised manually in BioEdit 7.0.5.3 (Hall 1999) and deposited at TreeBase (http://treebase.org/treebase-web/home.html, submission ID: 22474). Maximum Parsimony (MP), Bayesian Inference (BI) and Maximum Likelihood (ML) analyses were performed by using PAUP* 4.0b10 (Swofford 2002), MrBayes 3.1.2 (Ronquist and Huelsenbeck 2003) and RAxML 7.2.6 (Stamatakis 2006), respectively. The best models of evolution for BI were estimated by using MrModeltest 2.2 (Nylander 2004). The methods and parameter settings for the three kinds of phylogenetic analyses followed Liu et al. (2018).

Phylogeny results

The ITS-nrLSU sequences dataset contained 42 ITS and 53 nrLSU sequences from 53 samples representing 47 ingroup taxa and the outgroup (Table 1). Seven ITS and seven nrLSU sequences were generated for this study. The dataset had an aligned length of 2045 characters, of which 384 were parsimony informative. Maximum Parsimony (MP) analysis yielded 85 equally parsimonious trees. The best model estimated and applied in the Bayesian analysis was GTR+I+G. The average standard deviation of split frequencies of BI was 0.007863. ML and BI analyses resulted in almost the same tree topologies as that of MP analysis. Only the MP tree is shown in Fig. 1 with maximum likelihood and maximum parsimony bootstrap values ≥50% and BPP ≥0.95 labelled along the branches. In the tree, *A. bambusinus* and *A. isabellinus* were nested within the *A. cerussatus* (Bres.) Höhn. & Litsch. group (MP = 92%, BI = 1.00, ML = 87%). *Aleurodiscus subroseus* was clustered with *A. wakefieldiae*, but their relationship has no support in BI and ML analyses.

Taxonomy

Aleurodiscus bambusinus S.H. He & Y.C. Dai, sp. nov.

MycoBank: MB824755

Figs 2a-b, 3

Diagnosis. The species is distinct by having corticioid basidiomata, a compact texture, simple-septate generative hyphae, abundant acanthophyses, basidia with an acanthophysoid appendage and smooth basidiospores $7-10 \times 4-6 \mu m$ and growing on bamboo.

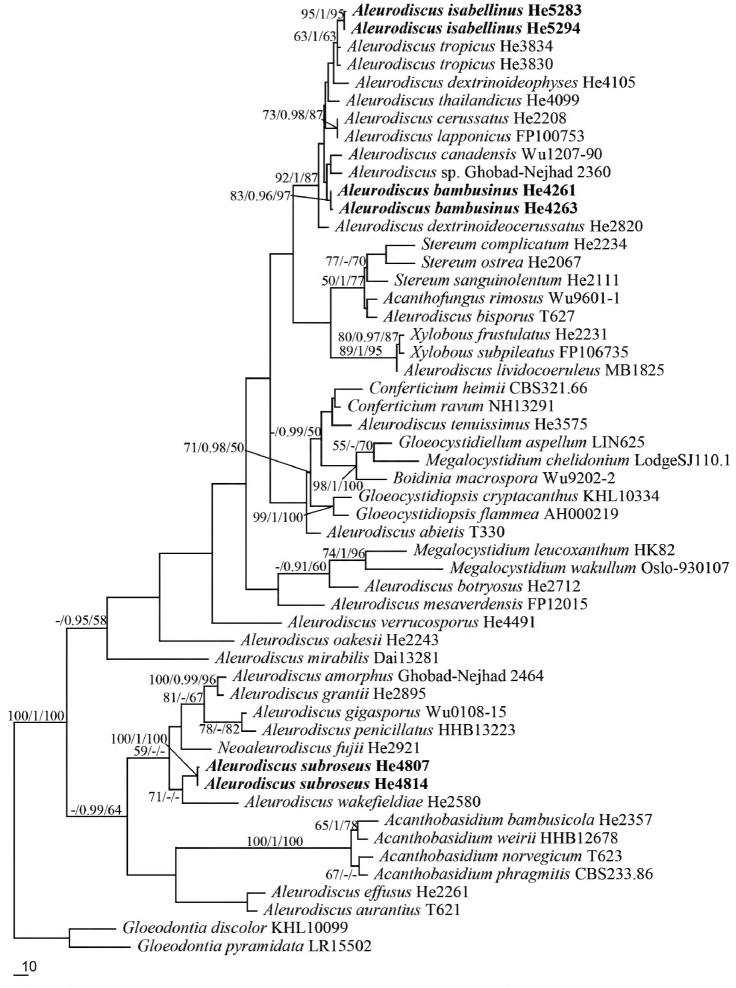


Figure 1. Maximum parsimony phylogeny of the combined ITS and nrLSU sequences data of Stereaceae. Branches are labelled with maximum parsimony and maximum likelihood bootstrap values $\geq 50\%$ and Bayesian posterior probabilities ≥ 0.95 (MP/BI/ML).

Table 1. Species and sequences used in the phylogenetic analyses. Newly generated sequences are set in bold.

Taxa	Voucher	Locality	ITS	nrLSU
Acanthobasidium bambusicola	He 2357	China	KU559343	KU574833
A. norvegicum	T 623	France	_	AY039328
A. phragmitis	CBS 233.86	France	_	AY039305
A. weirii	HHB 12678	USA	_	AY039322
Acanthofungus rimosus	Wu 9601-1	Taiwan	_	AY039333
Aleurodiscus abietis	Т 330	Canada	_	AY039324
A. amorphus	Ghobad-Nejhad 2464	China	KU559342	KU574832
A. aurantius	T 621	France	_	AY039317
A. bambusinus	He 4261	China	KY706207	KY706219
A. bambusinus	He 4263	China	KY706208	KY706218
A. bisporus	Т 627	Guadeloupe	_	AY039318
A. botryosus	He 2712	China	KX306877	KY450788
A. canadensis	Wu 1207-90	China	KY706203	KY706225
A. cerussatus	He 2208	China	KX306874	KY450785
A. dextrinoideocerussatus	He 2820	China	KY706206	MH109044
A. dextrinoideophyses	He 4105	China	MH109050	KY450784
A. effusus	He 2261	China	KU559344	KU574834
A. gigasporus	Wu 0108-15	China	KY706205	KY706213
A. grantii	He 2895	China	KU559347	KU574837
A. isabellinus	He 5283	China	MH109052	MH109046
A. isabellinus	He 5294	China	MH109053	MH109047
A. lapponicus	FP 100753	USA	_	AY039320
A. lividocoeruleus	MB 1825	USA	_	AY039314
A. mesaverdensis	FP 120155	USA	KU559359	KU574817
A. mirabilis	Dai 13281	China	KU559350	KU574839
A. oakesii	He 2243	USA	KU559352	KU574840
A. penicillatus	HHB 13223	USA	_	KU574816
A. sp.	Ghobad-Nejhad 2360	China	MH109051	MH109045
A. subroseus	He 4807	China	MH109054	MH109048
A. subroseus	He 4814	China	MH109055	MH109049
A. tenuissimus	He 3575	China	KX306880	KX842529
A. thailandicus	He 4099	Thailand	KY450781	KY450782
A. tropicus	He 3830	China	KX553875	KX578720
A. tropicus	He 3834	China	KX553876	KY706221
A. verrucosporus	He 4491	China	KY450786	KY450790
A. wakefieldiae	He 2580	China	KU559353	KU874841
Boidinia macrospora	Wu 9202-2	China: Taiwan	AF506377	AF506377
Conferticium heimii				
	CBS 321.66	Central African Republic	AF506381	AF506381
C. ravum	NH 13291	Republic Estonia	AF506381 AF506382	AF506381 AF506382
		Republic		
C. ravum	NH 13291 LIN 625 KHL 10334	Republic Estonia China: Taiwan Puerto Rico	AF506382 AF506432 AF506442	AF506382 AF506432 AF506442
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea	NH 13291 LIN 625 KHL 10334 AH 000219	Republic Estonia China: Taiwan Puerto Rico La Réunion	AF506382 AF506432 AF506442 AF506438	AF506382 AF506432 AF506442 AF506438
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico	AF506382 AF506432 AF506442 AF506438 AF506445	AF506382 AF506432 AF506442 AF506438 AF506445
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium M. leucoxanthum	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1 HK 82	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA Denmark	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium M. leucoxanthum M. wakullum	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1 HK 82 Oslo 930107	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA Denmark Tanzania	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420 AF506443	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420 AF506443
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium M. leucoxanthum M. wakullum Neoaleurodiscus fujii	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1 HK 82 Oslo 930107 He 2921	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA Denmark Tanzania China	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420 AF506443 KU559357	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420 AF506443 KU574845
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium M. leucoxanthum M. wakullum Neoaleurodiscus fujii Stereum complicatum	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1 HK 82 Oslo 930107 He 2921 He 2234	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA Denmark Tanzania China USA	AF506382 AF506432 AF506442 AF506445 AF506446 AF506441 AF506420 AF506443 KU559357 KU559368	AF506382 AF506432 AF506442 AF506445 AF506446 AF506441 AF506420 AF506443 KU574845 KU574828
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium M. leucoxanthum M. wakullum Neoaleurodiscus fujii Stereum complicatum S. ostrea	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1 HK 82 Oslo 930107 He 2921 He 2234 He 2067	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA Denmark Tanzania China USA USA USA	AF506382 AF506432 AF506442 AF506445 AF506445 AF506446 AF506441 AF506420 AF506443 KU559357 KU559368 KU559366	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420 AF506443 KU574845 KU574828
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium M. leucoxanthum M. wakullum Neoaleurodiscus fujii Stereum complicatum S. ostrea S. sanguinolentum	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1 HK 82 Oslo 930107 He 2921 He 2234 He 2067 He 2111	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA Denmark Tanzania China USA USA USA USA USA	AF506382 AF506432 AF506442 AF506445 AF506446 AF506441 AF506420 AF506443 KU559357 KU559368 KU559366 KU559367	AF506382 AF506432 AF506442 AF506445 AF506446 AF506441 AF506420 AF506443 KU574845 KU574828 KU574826 KU574827
C. ravum Gloeocystidiellum aspellum Gloeocystidiopsis cryptacanthus G. flammea Gloeodontia discolor G. pyramidata Megalocystidium chelidonium M. leucoxanthum M. wakullum Neoaleurodiscus fujii Stereum complicatum S. ostrea	NH 13291 LIN 625 KHL 10334 AH 000219 KHL 10099 LR 15502 LodgeSJ 110.1 HK 82 Oslo 930107 He 2921 He 2234 He 2067	Republic Estonia China: Taiwan Puerto Rico La Réunion Puerto Rico Columbia USA Denmark Tanzania China USA USA USA	AF506382 AF506432 AF506442 AF506445 AF506445 AF506446 AF506441 AF506420 AF506443 KU559357 KU559368 KU559366	AF506382 AF506432 AF506442 AF506438 AF506445 AF506446 AF506441 AF506420 AF506443 KU574845 KU574828

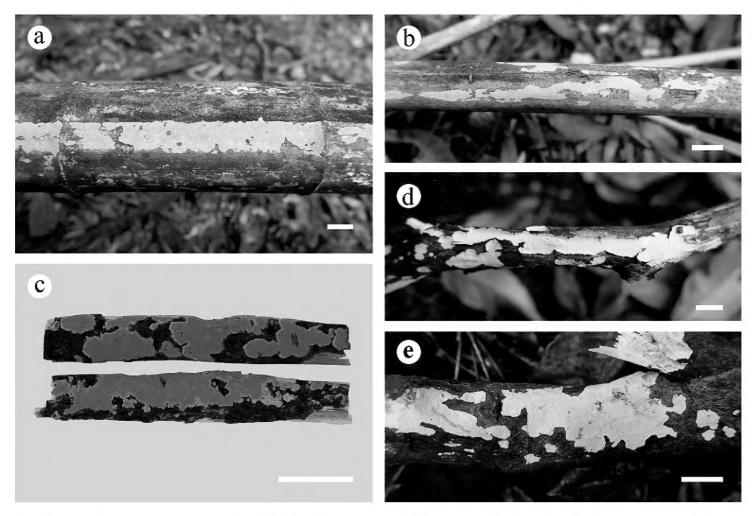


Figure 2. Basidiomata. **a–b** *Aleurodiscus bambusinus* (**a** He 4250 **b** holotype, He 4261) **c** *A. isabellinus* (holotype, KKN-2017-19) **d–e** *A. subroseus* (**d** He 5571 **e** He 4895). Scale bars: 1 cm.

Holotype. CHINA. Jiangxi Province, Yifeng County, Guanshan Nature Reserve, alt. ca. 800 m, on fallen culms and branches of bamboo, 10 Aug 2016, He 4261 (holotype, BJFC 023703).

Etymology. "Bambusinus" refers to the substrate of bamboo.

Basidiomata. Annual, resupinate, effused, closely adnate, inseparable from substrate, coriaceous, at first as small patches, later confluent up to 30 cm long and 2.5 cm wide, 180–300 μm thick. Hymenophore smooth, white (4A1) to yellowish-white (4A2) when young, becoming greyish-yellow [4B (3–4)] to brownish-orange [6C (5–8)] with age, uncracked or cracked with age; margin abrupt, indistinct, concolorous with hymenophore.

Microscopic structures. Hyphal system monomitic; generative hyphae simple-septate, colourless, thin- to thick-walled, scattered near the substrate, 2–4 μ m in diam. Subiculum thin to indistinct. Subhymenium thick, with compact texture, composed of acanthophyses and gloeocystidia. Acanthophyses abundant, hyphoid or distinctly swollen in the middle part, colourless, thin-walled, with abundant spines in apex, 30–40 \times 3–12 μ m. Gloeocystidia abundant, flexuous or slightly moniliform with one to several constrictions, slightly thick-walled, negative in sulphobenzaldehyde, 30–55 \times 8–13 μ m. Basidia subclavate to subcylindrical, colourless, slightly thick-walled, usually with a lateral acanthophysoid appendage, with four sterigmata and a basal simple septum, 25–35 \times 7–9 μ m. Basidiospores ellipsoid to broadly ellipsoid, bearing a dis-

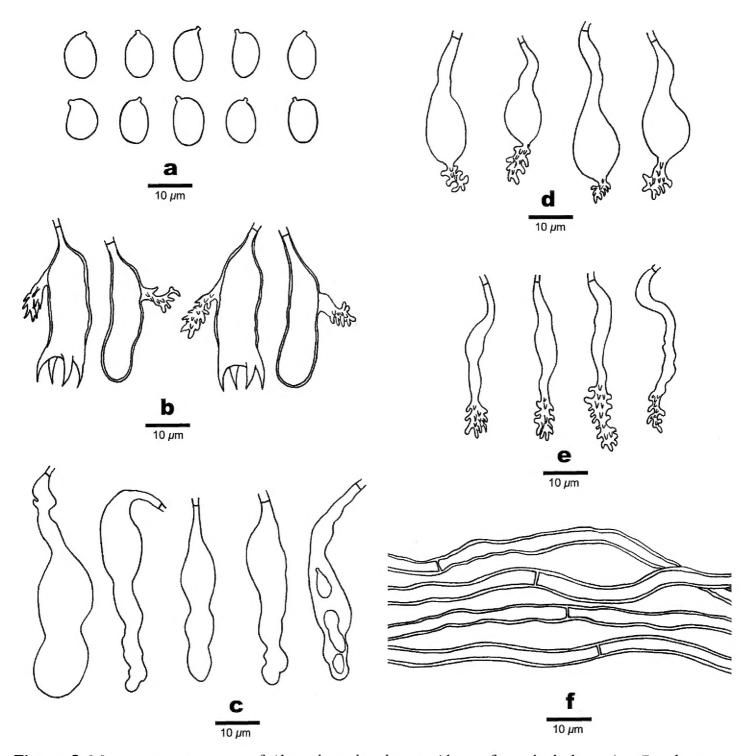


Figure 3. Microscopic structures of *Aleurodiscus bambusinus* (drawn from the holotype). **a** Basidiospores; **b** Basidia **c** Gloeocystidia **d–e** Acanthophyses **f** Generative hyphae.

tinct apiculus, colourless, thin-walled, smooth, amyloid, $7-10 \times 4-6$ µm, L = 8.7 µm, W = 4.9 µm, Q = 1.6–1.9 (n = 90/3).

Additional specimens examined. CHINA. Jiangxi Province, Yifeng County, Guanshan Nature Reserve, alt. ca. 800 m, on fallen culms and branches of bamboo, 10 Aug 2016, He 4250 (BJFC 023692) and He 4263 (BJFC 023705).

Remarks. Aleurodiscus bambusinus is morphologically similar and phylogenetically close to A. dextrinoideophyses S.H. He and A. tropicus L.D. Dai & S.H. He that also grow on bamboo in East Asia (Dai et al. 2017a, b). Aleurodiscus dextrinoideophyses differs from A. bambusinus by having apparently dextrinoid acanthophyses and smaller basidiospores (5–7 × 3–4 μ m, Dai et al. 2017b). Aleurodiscus tropicus differs from A. bambusinus by having a looser texture and slightly larger basidiospores (9–12 × 5–7.5 μ m, Dai et al. 2017a). The ITS similarity between A. bambusinus (He 4261)

and A. dextrinoideophyses (He 4105) is 95.6% of 434 base pairs and, between A. bambusinus (He 4261) and A. tropicus (He 3830), is 97.3% of 582 base pairs. Aleurodiscus aberrans G. Cunn. and A. rimulosus Núñez & Ryvarden are also similar to A. bambusinus, but they differ from this new species by having smooth basidia and growing on angiosperm wood outside of Asia (Núñez and Ryvarden 1997).

Aleurodiscus isabellinus S.H. He & Y.C. Dai, sp. nov.

MycoBank: MB824758

Figs 2c, 4

Diagnosis. The species is distinct by having soft, yellow to yellowish-brown and corticioid basidiomata, a loose texture, abundant yellow acanthophyses, simple-septate generative hyphae and smooth basidiospores $6-8.5 \times 3-4 \mu m$.

Holotype. CHINA. Yunnan Province, Dali County, Cangshan Nature Reserve, alt. ca. 2600 m, on fallen decorticated angiosperm branches, 27 Oct 2017, KKN-2017-19 (holotype in CFMR, isotype in BJFC).

Etymology. "Isabellinus" refers to the yellowish-brown basidiomata.

Basidiomata. Annual, resupinate, effused, adnate, inseparable from substrate, soft, membranaceous to coriaceous, at first as small patches, later confluent up to 15 cm long and 1 cm wide, 150–300 μ m thick. Hymenophore smooth, light orange [5A(4–5)], greyish-orange[5B(5–6)], orange [5B(7–8)] to brownish-yellow [5C(7–8)], uncracked or cracked with age; margin thinning out, fimbriate, white (5A1) when juvenile, becoming abrupt, indistinct, concolorous with hymenophore when mature.

Microscopic structures. Hyphal system monomitic, generative hyphae simple-septate, colourless, thin- to slightly thick-walled, straight, loosely interwoven, frequently branched and septate, 2–4 μm in diam. Acanthophyses abundant, colourless to yellow, thick-walled, hyphoid or arising laterally or apically from a clavate or cylindrical base $30–50 \times 5–7$ μm, with abundant spines in upper part, some hyphoid ones near substrate with long spines (branches) resembling binding hyphae. Gloeocystidia abundant, embedded, colourless, slightly thick-walled, subcylindrical or slightly moniliform, negative in sulphobenzaldehyde, $35–110 \times 5–8$ μm. Basidia clavate, colourless, thin-walled, with four sterigmata and a basal simple septum, $40–55 \times 6–7$ μm. Basidiospores ellipsoid to oblong ellipsoid, bearing a distinct apiculus, colourless, thin-walled, smooth, amyloid, (5.5–) 6–8.5 × (2.8–) 3–4 μm, L = 7 μm, W = 3.7 μm, Q = 1.9 (n = 24/1).

Additional specimens examined. CHINA. Yunnan Province, Dali County, Cangshan Nature Reserve, alt. ca. 2600 m, on small dead bamboo, 27 Oct 2017, He 5283 (BJFC 024801) and He 5287 (BJFC 024805); on fallen angiosperm branch, 27 Oct 2017, He 5294 (BJFC 024812); Jingdong County, Ailaoshan Nature Reserve, alt. 2450 m, on fallen angiosperm branch, 4 Oct 2017, C.L. Zhao 3843 (SWFC).

Remarks. All the studied specimens of *A. isabellinus* lack a true hymenium and only the holotype has a few basidia and basidiospores. *Aleurodiscus isabellinus* was nest-

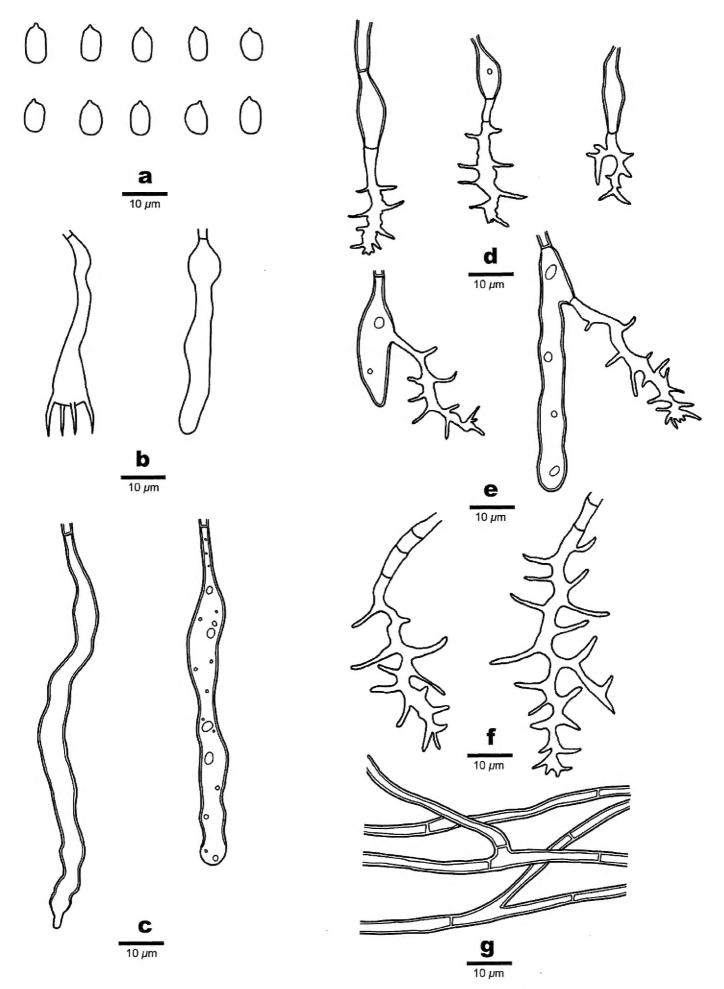


Figure 4. Microscopic structures of *Aleurodiscus isabellinus* (drawn from the isotype). **a** Basidiospores **b** A basidium and a basidiole **c** Gloeocystidia **d–f** Acanthophyses **g** Generative hyphae.

ed within the *A. cerussatus* group (Fig. 1). In this group, *Aleurodiscus thailandicus* S.H. He is similar to *A. isabellinus* by sharing the yellow basidiomata and acanthophyses, but differs by having two types of gloeocystida and acanthophyses without a clavate or

cylindrical base (Dai et al. 2017a). The ITS similarity between *A. isabellinus* (He 5283) and *A. thailandicus* (He 4099) is 93.6% of 578 base pairs. *Aleurodiscus thailandicus* was described from Thailand based on a fertile specimen on bamboo, but later several sterile specimens on bamboo from south-western China were identified as this species according to the sequence data. Morphologically, the soft and yellow to yellowish-brown basidiomata of *A. isabellinus* resemble the genus *Vararia* P. Karst. which belongs to Peniophoraceae according to phylogenetic analyses.

Aleurodiscus subroseus S.H. He & Y.C. Dai, sp. nov.

MycoBank: MB824757

Figs 2d-e, 5

Diagnosis. The species is distinct by having pinkish and corticioid basidiomata when fresh, clamped generative hyphae, moniliform gloeocystidia, presence of acanthophyses (acanthocystidia) and echinulate basidiospores $16-20 \times 11-14 \, \mu m$.

Holotype. CHINA. Guangxi Autonomous Region, Xing'an County, Mao'ershan Nature Reserve, alt. ca. 1600 m, on dead but still attached branch of living angiosperm tree, 13 Jul 2017, He 4807 (holotype, BJFC 024326).

Etymology. "Subroseus" (Lat.) refers to the pinkish basidiomata when fresh.

Basidiomata. Annual, resupinate, effused, closely adnate, inseparable from substrate, coriaceous, at first as small irregular patches, later confluent up to 35 cm long and 3 cm wide, up to 300 μm thick. Hymenophore smooth, pinkish-white (12A2), pink (12A3), pale orange (6A3) to light orange (6A4) when fresh, becoming pale orange (6A3), light orange [6A(4–5)], greyish-orange [6B(3–6)] to brownish-orange [6C(5–6)] when dry, uncracked; margin abrupt, white and distinct when fresh, becoming concolorous or darker than hymenophore and indistinct when dry, slightly elevated when mature.

Microscopic structures. Hyphal system monomitic, generative hyphae with clamp connections. Subiculum thin to indistinct. Subhymenium thickening with age, with embedded gloeocystidia, acanthophyses and crystals. Hyphae in this layer colourless, thin-walled, frequently branched and septate, agglutinated, 2–4 μm in diam. Gloeocystidia abundant, moniliform, with one to several constrictions, smooth, slightly thick-walled, negative in sulphobenzaldehyde, 45–70 × 6–12 μm. Acanthophyses (acanthocystidia) abundant, variable in shape and size, subclavate to subcylindrical, with few to many spines at apex, colourless, slightly thick-walled, 30–60 × 6–20 μm. Hyphidia scattered, thin-walled, colourless, rarely branched. Basidia clavate, slightly sinuous, colourless, thin-walled, smooth, with four sterigmata and a basal clamp connection, 52–80 × 13–17 μm. Basidiospores ellipsoid to broadly ellipsoid, bearing a distinct apiculus, colourless, slightly thick-walled, echinulate, strongly amyloid, 16–20 × 11–14 μm, L = 18.4 μm, W = 12.6 μm, Q = 1.5 (n = 90/3) (spines excluded).

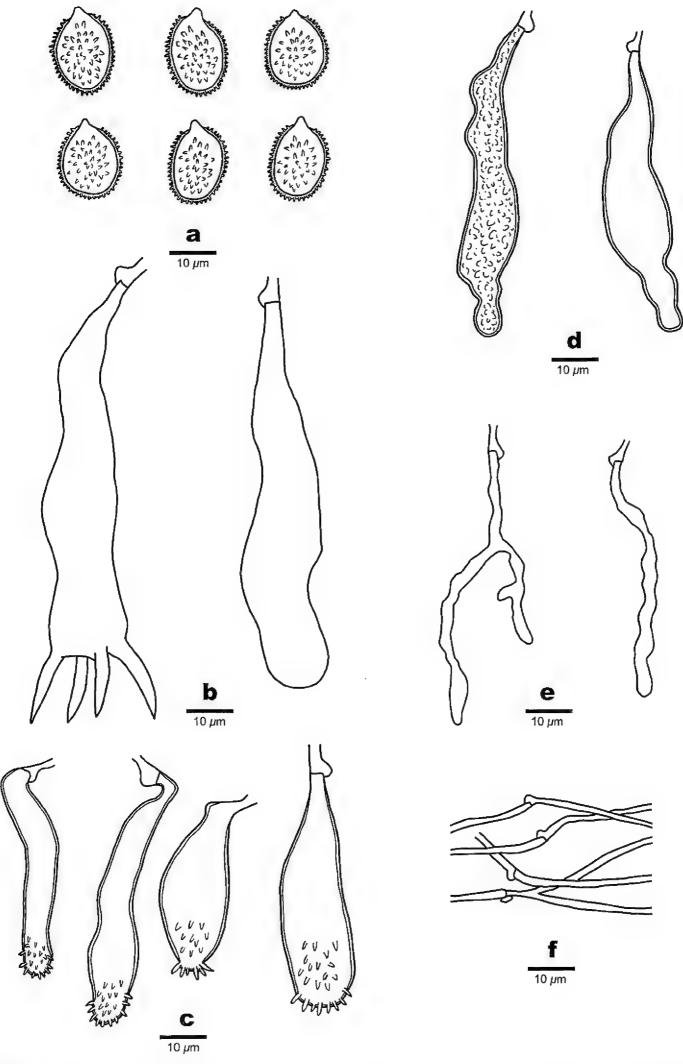


Figure 5. Microscopic structures of *Aleurodiscus subroseus* (drawn from the holotype). **a** Basidiospores; **b** A basidium and a basidiole **c** Acanthophyses **d** Gloeocystidia **e** Hyphidia **f** Generative hyphae.

Additional specimens examined. CHINA. Guangxi Autonomous Region, Xing'an County, Mao'ershan Nature Reserve, alt. ca. 1600 m, on dead but still attached branch of living angiosperm tree, 13 Jul 2017, He 4814 (BJFC 024333); Jinxiu County, Dayaoshan Nature Reserve, Yinshan Forest Park, alt. ca. 1500 m, on fallen angiosperm branch, 16 Jul 2017, He 4895 (BJFC 024414). Guizhou Province, Jiangkou County, Fanjingshan Nature Reserve, alt. 1500–2000 m, on dead but still attached branch of living angiosperm tree, 11 Jul 2018, He 5558 (BJFC); 12 Jul 2018, He 5571, He 5577, He 5581, He 5585, He 5589 and He 5593 (BJFC).

Remarks. Aleurodiscus subroseus is morphologically similar and phylogenetically close to A. wakefieldiae Boidin & Beller (Fig. 1), but the latter differs by having longer basidia (80–180 μm) and larger basidiospores (20–28 × 14–20 μm, Núñez and Ryvarden 1997). Aleurodiscus penicillatus Burt is similar to A. subroseus, but differs by growing on gymnosperm wood and having wider basidiospores (13–17 μm, Núñez and Ryvarden 1997). Aleurodiscus mirabilis (Berk. & M.A. Curtis) Höhn. also has pinkish fresh basidiomata and is widely distributed in southern China. However, it can be easily distinguished from A. subroseus by having basally warted basidia and larger basidiospores (24–28 × 14–17 μm, Núñez and Ryvarden 1997). In the phylogenetic tree (Fig. 1), A. penicillatus and A. mirabilis are distantly related to A. subroseus. Aleurodiscus corticola Gorjón et al. from Argentina on bark of living Nothofagus dombeyi also has moniliform gloeocystidia and similar basidiospores with A. subroseus, but differs by having pulvinate and tuberculate basidiomata and absence of acanthophyses (Gorjón et al. 2013).

Key to 26 species of Aleurodiscus s.l. in China

Acanthobasidium Oberw., Aleurocystidiellum P.A. Lemke and Neoaleurodiscus Sheng H. Wu are used for some species. Basidiospores data are from Núñez & Ryvarden (1997) or otherwise measured by the authors.

Basidiospores smooth2	1
Basidiospores ornamented11	_
Acanthophyses absent3	2
Acanthophyses present4	_
Basidiospores thick-walled, 23–27 × 16–21 μm; on <i>Rhododendron</i>	3
Neoaleurodiscus fujii	
Basidiospores thin-walled, $18-23 \times 14-19 \mu m$; on QuercusA. ljubarskii	_
Basidia with two sterigmata; basidiospores >12 μm long	4
Basidia with four sterigmata; basidiospores <12 µm long	_
Generative hyphae simple-septate6	5
Generative hyphae clamped10	_
Acanthophyses apparently dextrinoid	6
Acanthophyses indextrinoid7	_

7	Basidia smooth; acanthophyses yellow
_	Basidia with an acanthophysoid appendage; acanthophyses colourless9
8	Gloeocystidia of two types; acanthophyses hyphoid
_	Gloeocystidia of one type; acanthophyses hyphoid, subclavate to subcy-
	lindrical
9	Texture loose; basidiospores $9-12 \times 5-7.5 \mu m$
_	Texture compact; basidiospores $7-10 \times 4-6 \mu m$
10	Acanthophyses apparently dextrinoid
_	Acanthophyses indextrinoid
11	Acanthophyses absent
_	Acanthophyses present
12	Generative hyphae simple-septate
_	Generative hyphae clamped
13	Basidiomata discoid; basidiospores >20 µm long
_	Basidiomata corticioid; basidiospores <20 µm long14
14	Basidiospores <8 µm long
_	Basidiospores >8 μm long
15	Basidiospores $12-17 \times 10-15 \mu m$; on angiosperm wood A. ryvardenii
_	Basidiospores $8-11.5 \times 6-8.5 \mu m$; on bamboo
16	Basidiospores >20 µm long
_	Basidiospores <20 µm long
17	On Quercus
_	On gymnosperm
18	Encrusted skeletocystidia present; on Abies
_	Moniliform gloeocystidia present; on PinusAleurocystidiellum tsugae
19	Acanthophyses amyloid
_	Acanthophyses non-amyloid
20	Basidiospores globose; on bamboo
_	Basidiospores ellipsoid; on wood
21	On gymnosperm22
_	On angiosperm23
22	Basidiospores $16-21 \times 12-17 \mu \text{m}$
_	Basidiospores $26-38 \times 20-28 \ \mu \text{m}$
23	Basidiomata white when fresh; acanthophyses rare
_	Basidiomata pinkish when fresh; acanthophyses abundant24
24	Basidiospores $16-20 \times 11-14 \mu \text{m}$
_	Basidiospores >20 μm long, >14 μm wide
25	Acanthophyses hyphoid, covered with spines at whole upper part; basidia
	and gloeocystidia covered with spines at basal part; basidiospores usually
	D-shaped
_	Acanthophyses hyphoid to clavate, covered with spines only at apex; basidia
	and gloeocystidia smooth; basidiospores ellipsoid
	o ,

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References

- Dai LD, He SH (2016) New species and new records of *Aleurodiscus* s.l. (Basidiomycota) in China. Mycological Progress 15: 717–730. https://doi.org/10.1007/s11557-016-1202-z
- Dai LD, He SH (2017) A new species and a new combination of *Aleurodiscus* s.l. (Russulales, Basidiomycota). Mycosphere 8: 908–916. https://doi.org/10.5943/mycosphere/8/7/7
- Dai LD, Wu SH, Nakasone KK, Burdsall Jr HH, He SH (2017a) Two new species of *Aleuro-discus* s.l. (Russulales, Basidiomycota) on bamboo from tropics. Mycoscience 58: 213–220. https://doi.org/10.1016/j.myc.2017.02.001
- Dai LD, Zhao Y, He SH (2017b) Three new species of *Aleurodiscus* s.l. (Russulales, Basidiomycota) on bamboos from East Asia. Cryptogamie Mycologie 38: 227–239. https://doi.org/10.7872/crym/v38.iss2.2017.227
- Gorjón SP, Greslebin AG, Rajchenberg M (2013) The genus *Aleurodiscus* s.l. (Stereaceae, Russulales) in the Patagonian Andes. Mycological Progress 12: 91–108. https://doi.org/10.1007/s11557-012-0820-3
- Hall TA (1999) Bioedit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95–98.
- Kornerup A, Wanscher JH (1978) Methuen handbook of colour (3rd edn). Eyre Methuen, London, 252 pp.
- Katoh K, Toh H (2008) Recent developments in the MAFFT multiple sequence alignment program. Briefings in Bioinformatics 9: 286–298. https://doi.org/10.1093/bib/bbn013
- Larsson KH (2007) Re-thinking the classification of corticioid fungi. Mycological Research 111: 1040–1063. https://doi.org/10.1016/j.mycres.2007.08.001
- Liu SL, Nakasone KK, Wu SH, He SH, Dai YC (2018) Taxonomy and phylogeny of *Lopharia* s.s., *Dendrodontia*, *Dentocorticium* and *Fuscocerrena* (Basidiomycota, Polyporales). MycoKeys 32: 25–48. https://doi.org/10.3897/mycokeys.32.23641
- Núñez M, Ryvarden L (1997) The genus *Aleurodiscus* (Basidiomycotina). Synopsis Fungorum 12: 1–164
- Nylander JAA (2004) MrModeltest 2.2. Program distributed by the author. Evolutionary Biology Centre, Uppsala University, Uppsala.
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19: 1572–1574. https://doi.org/10.1093/bioinformatics/btg180

- Stamatakis A (2006) RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics 22: 2688–2690. https://doi.org/10.1093/bioinformatics/btl446
- Swofford DL (2002) PAUP*: Phylogenetic analysis using parsimony (*and other methods). Version 4.0b10. Sinauer Associates, Sunderland, Massachusetts.
- White TJ, Bruns TD, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR protocols, a guide to methods and applications. Academic, San Diego, 315–322. https://doi.org/10.1016/b978-0-12-372180-8.50042-1
- Wu SH, Hibbett DS, Binder M (2001) Phylogenetic analyses of *Aleurodiscus* s.l. and allied genera. Mycologia 93: 720–731. https://doi.org/10.2307/3761826